

## Fusion BioLabs Short Protocol for Transformation of Phage Display Cells

### 1. Product description

Catalogue #	Strain	Amber Suppressor (Y/N)	Main application	Genotype
PDCE	ER2738	N	Phage display library construction	<i>F'</i> proA+B+ lacIq Δ(lacZ)M15 zzf::Tn10 (tet) <i>r</i> ] fhuA2 glnVΔ(lac-proAB) thi-1Δ(hsdS-mcrB)5
PDCS	SS320	Y	Antibody fragment expression	<i>[F'</i> proA+B+ lacIqlacZΔM15 Tn10 (tet) <i>r</i> ] hsdR mcrB araD139 Δ(araABC-leu)7679 ΔlacX74 galUgalK rpsL thi
PDCT	TG1	N	Phage display library construction	<i>[F'</i> traD36 proA+B+ lacIqZ ΔM15] supE thi-1 Δ(lac-proAB) Δ(mcrB-hsdSM)5(rK- mK-)

### 2. Panning Protocol using biotin labelled antigen target

- 1) Take Recovery Medium and 1.5 ml or 2.0 ml sterilized microcentrifuge tube readily available at room temperature (one tube for each transformation reaction).
- 2) Place the microcentrifuge tubes on ice (one microfuge tube for each transformation reaction).
- 3) Remove Phage Display Cells from the -80 °C freezer and place on wet ice until they thaw completely (10-15 minutes).
- 4) When the cells are thawed, mix them by tapping gently. Aliquot 50 µL of cells into the chilled microcentrifuge tubes on ice.
- 5) Add 1 µL monoclonal phage ELISA validated and/or Sequencing confirmed candidate (from panning of scFv, Fab, VHH, affibody or peptide phage library) to the 50 µL of cells on ice for 10 min. Stir briefly with pipette tip; do not pipet up and down to mix, which can introduce air bubbles and warm the cells.
- 6) Heat shock at 42°C water bath for 30 seconds
- 7) Put the heat shocked microfuge tube on ice immediately; add 450 µL of Recovery Medium to the microfuge tube.
- 8) Place the tube in a shaking incubator at 250 rpm for 1 hour at 37 °C.
- 9) Spread up to 100 µL of transformed cells on 2xYT agar plate containing the appropriate antibiotic.
- 10) Incubate the plates overnight at 37 °C.

11) Transformed single clone can be further grown in 2xYT medium to isolate for down-stream application, such as DNA plasmid preparation, or in TB medium for antibody fragment expression.

### 3. Media recipes

#### 3.1 2xYT Agar plates for plate transformed cells (per liter)

16 g tryptone

10 g yeast extract

5 g NaCl

15 g agar

*Add all components to distilled water. Adjust pH to 7.0 with 5N NaOH. Autoclave and cool to 55 °C.*

#### 3.2 2xYT Medium for growth of transformants (per liter)

16 g tryptone

10 g yeast extract

5 g NaCl

15 g agar

*Add all components to distilled water. Adjust pH to 7.0 with 5N NaOH. Autoclave and cool to 55 °C.*

#### 3.3 TB media for antibody fragment expression (per liter)

12 g tryptone

24 g yeast extract

12.5 g  $K_2HPO_4$

2.3 g  $KH_2PO_4$

4 ml 100% glycerol

*Add all components to deionised water. Autoclave and cool to 55 °C.*