

Fusion BioLabs Short Protocol for Antibody Expression Vector Kit

1. Product Description

	SKU	Heavy chain vector	Description	SKU	Light chain vector	Description
Human	AEV-01	pFB-CHIg-hG1	Human IgG1 Mammalian Expression Vector	AEV-09	pFB-CLIg-hk	Human Igk Mammalian Expression Vector
	AEV-02	pFB-CHIg-hG2	Human IgG2 Mammalian Expression Vector	AEV-10	pFB-CLIg-hl	Human Igλ2 Mammalian Expression Vector
	AEV-03	pFB-CHIg-hG3	Human IgG3 Mammalian Expression Vector			
	AEV-04	pFB-CHIg-hG4	Human IgG4 Mammalian Expression Vector			
	AEV-05	pFB-CHIg-hA1	Human IgA Mammalian Expression Vector			
	AEV-21	pFB-CHIg-hA2	Human IgA Mammalian Expression Vector			
	AEV-06	pFB-CHIg-hD	Human IgD Mammalian Expression Vector			
	AEV-07	pFB-CHIg-hE	Human IgE Mammalian Expression Vector			
	AEV-08	pFB-CHIg-hM (pentamer)	Human IgM Mammalian Expression Vector			
	AEV-20	pFB-CHIg-hM (monomer)	Human IgM Mammalian Expression Vector			
Mouse	AEV-11	pFB-CHIg-mG1	Mouse IgG1 Mammalian Expression Vector	AEV-19	pFB-CLIg-mk	Mouse Igk Mammalian Expression Vector
	AEV-12	pFB-CHIg-mG2a	Mouse IgG2a Mammalian	AEV-23	pFB-CLIg-mI1	Mouse Igλ1 Mammalian

			Expression Vector			Expression Vector
	AEV-13	pFB-CHIg-mG2b	Mouse IgG2b Mammalian Expression Vector	AEV-24	pFB-CLIg-mI2	Mouse Igλ2 Mammalian Expression Vector
	AEV-14	pFB-CHIg-mG3	Mouse IgG3 Mammalian Expression Vector			
	AEV-15	pFB-CHIg-mA	Mouse IgA Mammalian Expression Vector			
	AEV-16	pFB-CHIg-mD	Mouse IgD Mammalian Expression Vector			
	AEV-22	pFB-CHIg-mE	Mouse IgE Mammalian Expression Vector			
	AEV-17	pFB-CHIg-mM (pentamer)	Mouse IgM Mammalian Expression Vector			
	AEV-18	pFB-CHIg-mM (monomer)	Mouse IgM Mammalian Expression Vector			
Chicken	AEV-28	pFB-CHIg-cY	Chicken IgY Mammalian Expression Vector	AEV-29	pFB-CLIg-cl	Chicken Igλ Mammalian Expression Vector
Rabbit	AEV-25	pFB-CHIg-rG	Rabbit IgG Mammalian Expression Vector	AEV-26	pFB-CLIg-rk1	Rabbit Igk Mammalian Expression Vector
				AEV-27	pFB-CLIg-rl	Rabbit Igλ Mammalian Expression Vector
Monkey	AEV-30	pFB-CHIg-rmG	Rhesus monkey IgG Mammalian Expression Vector	AEV-31	pFB-CLIg-rmk	Rhesus monkey Igk Mammalian Expression Vector

2. PROTOCOL

2.1 Obtaining VH and VL sequences

You could obtain VH and VL sequences from either **gene synthesis** or **PCR amplification** from your template:

For gene synthesis, a 5'-end with sequence (5'-TAGTAGCAACTGCA**ACCGGT**GTACATTCA-3') and 3'-end with the following sequence (different, see table below) should be appended to your VH or VL (Vk or Vλ) ends.

Note: There is no need to add signal sequence to your VH and VL fragment.

hlgG1	5'-TAGTAGCAACTGCA ACCGGT GTACATTCA-VH-GT CTCGAG Cgcctccaccaagggc-3'
hlgG2	5'-TAGTAGCAACTGCA ACCGGT GTACATTCA-VH-GT CTCGAG Cgcctccaccaagggc-3'
hlgG3	5'-TAGTAGCAACTGCA ACCGGT GTACATTCA-VH-GT CTCGAG Cgctagcaccaagggc-3'
hlgG4	5'-TAGTAGCAACTGCA ACCGGT GTACATTCA-VH-GT CTCGAG Cgcctccaccaagggc-3'
hlgA1	5'-TAGTAGCAACTGCA ACCGGT GTACATTCA-VH-GT CTCGAG Cgcatccccgaccagc-3'
hlgA2	5'-TAGTAGCAACTGCA ACCGGT GTACATTCA-VH-GT CTCGAG Cgcatccccgaccagc-3'
hlgD	5'-TAGTAGCAACTGCA ACCGGT GTACATTCA-VH-GT CTCGAG Cgcacccaccaagggc-3'
hlgE	5'-TAGTAGCAACTGCA ACCGGT GTACATTCA-VH-GT CTCGAG Cgcctccacacagagc-3'
hlgM	5'-TAGTAGCAACTGCA ACCGGT GTACATTCA-VH-GT CTCGAG Cgggagtgcacccg-3'
hlgk	5'-TAGTAGCAACTGCA ACCGGT GTACATTCA-Vk-GT CTCGAG Cgaactgtggctgcac-3'
hlgλ	5'-TAGTAGCAACTGCA ACCGGT GTACATTCA-Vλ-TTG CTCGAG ggtcagcccaagggc-3'
mlgG1	5'-TAGTAGCAACTGCA ACCGGT GTACATTCA-VH-GT CTCGAG Cgcaaaaacgacacc-3'
mlgG2a	5'-TAGTAGCAACTGCA ACCGGT GTACATTCA-VH-GT CTCGAG Cgcaaaaacaacagcc-3'
mlgG2b	5'-TAGTAGCAACTGCA ACCGGT GTACATTCA-VH-GT CTCGAG Cgcaaaaacaacacc-3'
mlgG3	5'-TAGTAGCAACTGCA ACCGGT GTACATTCA-VH-GT CTCGAG Cacaacaacagccca-3'
mlgA	5'-TAGTAGCAACTGCA ACCGGT GTACATTCA-VH-GT CTCGAG Cgagtctgcgagaaat-3'
mlgD	5'-TAGTAGCAACTGCA ACCGGT GTACATTCA-VH-GT CTCGAG Cgataaaaaggaacct-3'
mlgE	5'-TAGTAGCAACTGCA ACCGGT GTACATTCA-VH-GT CTCGAG Cgctagcatcaggaac-3'
mlgM	5'-TAGTAGCAACTGCA ACCGGT GTACATTCA-VH-GT CTCGAG Cagtcagtccttcca-3'
mlgk	5'-TAGTAGCAACTGCA ACCGGT GTACATTCA-Vk-CGT CTCGAG cgggctgatgctgca-3'
mlgλ1	5'-TAGTAGCAACTGCA ACCGGT GTACATTCA-Vλ-GT CTCGAG Cggccagcccaagtct-3'
mlgλ2	5'-TAGTAGCAACTGCA ACCGGT GTACATTCA-Vλ-GT CTCGAG Cggtcagcccaagtcc-3'

rlgG	5'-TAGTAGCAACTGCAACCGGTGTACATTCA-VH-GTCTCGAGCgggcaacctaaaggct-3'
rlgk	5'-TAGTAGCAACTGCAACCGGTGTACATTCA-Vk-GTCTCGAGCggtgatccagttgca-3'
rlgl	5'-TAGTAGCAACTGCAACCGGTGTACATTCA-Vλ-GTCTCGAGCcagcccgcggtgacc-3'
rmlgG	5'-TAGTAGCAACTGCAACCGGTGTACATTCA-VH-GTCTCGAGCgcctccaccaagggc-3'
rmlgk	5'-TAGTAGCAACTGCAACCGGTGTACATTCA-Vk-GTCTCGAGCcgagctgtggctgca-3'
clgY	5'-TAGTAGCAACTGCAACCGGTGTACATTCA-VH-GTCTCGAGCgcctccccacctcc-3'
clgl	5'-TAGTAGCAACTGCAACCGGTGTACATTCA-Vλ-GTCTCGAGCgccccaccatcacc-3'

For PCR amplification, the Forward Primer and Reverse Primer should be as following. The optimized annealing temperature should be 53-58°C. For best in-frame insert, the resulting amplicons must be sequenced before or after the cloning into the expression vector.

Forward sequencing primer (pCMV5F): 5'-ATGGGCGGTAGGCGTGTA-3' (included in the Kit).

Note: Forward Primer's N(12-18) is from your VH or VL coding region (no need adding signal peptide sequence); Reverse Primer's N(12-18) is the terminal coding sequence of your VH or VH.

	Forward Primer	Reverse Primer
hlgG1	5'-TAGTAGCAACTGCAACCGGTGTACATTCAN (12-18) -3'	5'-gcccttggtggagggcGCTCGAGACN (12-18) -3'
hlgG2	5'-TAGTAGCAACTGCAACCGGTGTACATTCAN (12-18) -3'	5'-gcccttggtggagggcGCTCGAGACN (12-18) -3'
hlgG3	5'-TAGTAGCAACTGCAACCGGTGTACATTCAN (12-18) -3'	5'-gcccttggtgctagcGCTCGAGACN (12-18) -3'
hlgG4	5'-TAGTAGCAACTGCAACCGGTGTACATTCAN (12-18) -3'	5'-gcccttggtggagggcGCTCGAGACN (12-18) -3'
hlgA1	5'-TAGTAGCAACTGCAACCGGTGTACATTCAN (12-18) -3'	5'-gctggtcggggatgGCTCGAGACN (12-18) -3'
hlgA2	5'-TAGTAGCAACTGCAACCGGTGTACATTCAN (12-18) -3'	5'-gctggtcggggatgGCTCGAGACN (12-18) -3'
hlgD	5'-TAGTAGCAACTGCAACCGGTGTACATTCAN (12-18) -3'	5'-agccttggtgggtgGCTCGAGACN (12-18) -3'
hlgE	5'-TAGTAGCAACTGCAACCGGTGTACATTCAN (12-18) -3'	5'-gctctgtgtggagggcGCTCGAGACN (12-18) -3'



hlgM	5'-TAGTAGCAACTGCAACCGGTGTACATTCAN (12-18) -3'	5'-ggcggatgcactcccGCTCGAGACN (12-18) -3'
hlgk	5'-TAGTAGCAACTGCAACCGGTGTACATTCAN (12-18) -3'	5'-gtgcagccacagttcGCTCGAGACN (12-18) -3'
higl	5'-TAGTAGCAACTGCAACCGGTGTACATTCAN (12-18) -3'	5'-agccttgggctgaccGCTCGAGCAAN (12-18) -3'
mlgG1	5'-TAGTAGCAACTGCAACCGGTGTACATTCAN (12-18) -3'	5'-gggtgtcgttttggcGCTCGAGACN (12-18) -3'
mlgG2a	5'-TAGTAGCAACTGCAACCGGTGTACATTCAN (12-18) -3'	5'-ggctgttgttttggcGCTCGAGACN (12-18) -3'
mlgG2b	5'-TAGTAGCAACTGCAACCGGTGTACATTCAN (12-18) -3'	5'-gggtgttgttttggcGCTCGAGACN (12-18) -3'
mlgG3	5'-TAGTAGCAACTGCAACCGGTGTACATTCAN (12-18) -3'	5'-tggggctgttgttGCTCGAGACN (12-18) -3'
mlgA	5'-TAGTAGCAACTGCAACCGGTGTACATTCAN (12-18) -3'	5'-atctctcgagactcGCTCGAGACN (12-18) -3'
mlgD	5'-TAGTAGCAACTGCAACCGGTGTACATTCAN (12-18) -3'	5'-aggttcctttttatcGCTCGAGACN (12-18) -3'
mlgE	5'-TAGTAGCAACTGCAACCGGTGTACATTCAN (12-18) -3'	5'-gttcctgatgctagcGCTCGAGACN (12-18) -3'
mlgM	5'-TAGTAGCAACTGCAACCGGTGTACATTCAN (12-18) -3'	5'-tgggaaggactgactGCTCGAGACN (12-18) -3'
mlgk	5'-TAGTAGCAACTGCAACCGGTGTACATTCAN (12-18) -3'	5'-tgcagcatcagcccGCTCGAGACGN (12-18) -3'
mlgl1	5'-TAGTAGCAACTGCAACCGGTGTACATTCAN (12-18) -3'	5'-agacttgggctggccGCTCGAGACN (12-18) -3'
mlgl2	5'-TAGTAGCAACTGCAACCGGTGTACATTCAN (12-18) -3'	5'-ggacttgggctgaccGCTCGAGACN (12-18) -3'
rlgG	5'-TAGTAGCAACTGCAACCGGTGTACATTCAN (12-18) -3'	5'-agccttaggttgcccGCTCGAGACN (12-18) -3'
rlgk	5'-TAGTAGCAACTGCAACCGGTGTACATTCAN (12-18) -3'	5'-tgcaactggatcaccGCTCGAGACN (12-18) -3'
rlgl	5'-TAGTAGCAACTGCAACCGGTGTACATTCAN (12-18) -3'	5'-ggtcaccgctgggctgGCTCGAGACN (12-18) -3'
rmlgG	5'-TAGTAGCAACTGCAACCGGTGTACATTCAN (12-18) -3'	5'-gcccttggaggaggcGCTCGAGACN (12-18) -3'
rmlgk	5'-TAGTAGCAACTGCAACCGGTGTACATTCAN (12-18) -3'	5'-tgcagccacagctcgGCTCGAGACN (12-18) -3'
clgY	5'-TAGTAGCAACTGCAACCGGTGTACATTCAN (12-18) -3'	5'-ggaggtgggggaggcGCTCGAGACN (12-18) -3'
clgl	5'-TAGTAGCAACTGCAACCGGTGTACATTCAN (12-18) -3'	5'-ggtgatggtgggggcGCTCGAGACN (12-18) -3'

2.2 Cloning into pFB-CHlg (heavy chain expression vector) and pFB-CLlg (light chain expression vector)

Once the VH and VL sequences have been obtained, the VH and VL could be cloned into the pFB-CHlg heavy chain expression vector, and the pFB-CLlg light chain expression vector, respectively. Two methods are available:

Restriction Enzyme Cloning

There is 5'-end AgeI and 3'-end XhoI for all pFB-CHIg and pFB-CLIg expression vector. All of our antibody expression vector are compatible with high throughput platform.

1) Digestion setup

Component	50 μ l reaction
VH or VL Inserts / pFB-CHIg or pFB-CLIg vector)	1 μ g / 5 μ g
Restriction buffer (10x)	5 μ l (1x)
AgeI	5 units
XhoI	20 units
Nuclease-free H ₂ O	to 50 μ l

- Incubate at 37°C for 1-3 hours.
- Run agarose gel to purify the digested inserts and vector backbone.

2) T4 DNA ligation

Component	20 μ l reaction
T4 DNA ligation buffer (10x)	2 μ l
Vector DNA	80 ng
Insert DNA	15 ng
T4 DNA ligase	400 units
Nuclease-free H ₂ O	to 20 μ l

- Mix gently and microfuge briefly, and incubate at 16°C or 4°C overnight or room temperature for 30 min.
- Transformation: chill on ice and transform 5 μ l of the reaction into 50 μ l competent cells.

Cloning through homologous assembly

There are many convenient kits for this method from different supply. We recommend NEBuilderHiFi DNA Assembly Kit (Cat# E2621S).

Component	5 μ l reaction in PCR tube
Vector DNA	45 ng
Insert DNA	4.5 ng
NEBuilder HiFi DNA Assembly Master Mix	2.5 μ l
Nuclease-free H ₂ O	to 5 μ l

- Mix gently and microfuge briefly, move the PCR tube to previously set PCR program: 50°C, 15 minutes, 4°C, 5 minutes.
- Store PCR reaction tube on ice or at -20°C for subsequent transformation.
- Transformation: chill on ice and transform 2.5 μ l of the reaction into 25 μ l competent cells.

2.3 Antibody Production

Cotransfect mammalian cells, such as CHO and 293 cells, with the sequencing confirmed expression plasmid pair, pFB-CHIg encoding the heavy chain, and pFB-CLLg encoding the light chain. Typically, we recommend using a ratio of 2:3 of pFB-CHIg: pFB-CLLg plasmids.

Note: Antibody production after transfection, you may take an aliquot of growth medium and perform SDS-PAGE, target protein-specific ELISA, or bioassay of choice to determine that your cells are producing your antibody of interest.

2.4 Antibody Purification

The resulting IgG, IgA, IgD, IgE, and IgM antibody can be purified from the CHO supernatant or HEK293 supernatant using the appreciate Protein A, Protein G, Protein L or antigen-coupled resin for affinity chromatography.